

SYNTHESIS AND EVALUATION OF VARIOUS PHYSICO-CHEMICAL PARAMETER OF NOVEL 1,3,5-TRIAZINES DERIVATIVES AS POTENT ANTIMICROBIAL AGENTS

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ABSTRACT: - Heterocyclic chemistry offers an example for the lack of distinct demarcations; in fact, it pervades the plurality of the other chemical disciplines. Heterocycles are inextricably woven into the life processes. The vital interest of the pharmaceutical and agrochemical industries in heterocycles is often connected with their natural occurrence. A series of 1, 3, 5-triazines derivatives were designed, synthesized and evaluated their biological activity. The preliminary investigation showed that most compounds displayed well to excellent potency against selected micro organism. The structure of the novel compounds was elucidated on the basis of IR, ¹H-NMR and elemental analysis.

KEYWORDS: s-triazine, lipophilicity, antibacterial and antifungal activity.

INTRODUCTION

A wide range of 1,3,5-triazines exhibit selective herbicidal properties, Simazine and atrazines are the organic compounds containing striazine skeleton are most important herbicides.[1] Triazines derivative Altretamine and triethylene melamine (TEM) shown activity against leukemia. Compounds contain 1,3,5-triazine as a lead moiety possessing a wide spectrum of biological activities such as anti-cancer[2-6], antiviral [7], bactericidal [8-10], fungicidal[11-12], antimalarial agents[13-14] and antituberculosis [15]. In addition, the interest in 1,3,4-oxadiazole also one of important heterocyclic compound show significant biological activity such as antibacterial and fungicidal.[16-18] Considerable attention has been paid to the analysis of chemical in the 1,3 thiazole moiety , due to their widespread use in medicinal chemistry and their subsequent degradation in biological systems. For initial chemical screening of the activity of newly synthesized compounds it is recommended first to determine their lipophilicity, an important physico-chemical property in relation to biological activity. Lipophilicity is difficult to quantitative, but the most widely accepted measure of lipophilicity is the octanol-water partition coefficient,

defined as the ratio of the concentrations of the solute in the two phases of a saturated 1-octanol-water system. Measurement of the octanol-water partition coefficient is achieved by an alternative method, reversed-phase liquid chromatography. Reversed-phase thin-layer chromatography (RP TLC) has been found to offer a rapid method for the analysis of a large number of substituted 1,3 thiazole type compounds.

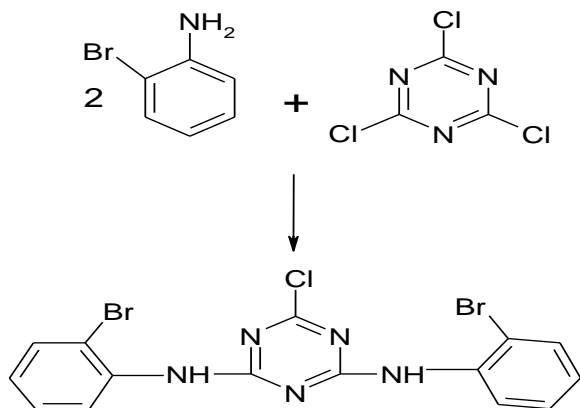
MATERIAL AND METHODS

All chemicals and solvents procured were of analytical grade and used directly without further purification. All melting points were determined in PMP-DM scientific melting point apparatus and are uncorrected. Thin-layer chromatography (TLC) was performed using silica gel-G coated Al-plates (0.5 mm thickness, Merck, Germany) using appropriate mobile phase system and spots were visualized under UV and Iodine chamber. ¹H NMR spectra were recorded on a Bruker Avance-II 400 MHz spectrophotometer (Bruker Bioscience, USA) using DMSO as a solvent and TMS as internal standard (chemical shifts in δ ppm). Infra Red spectra were recorded on FT Shimadzu spectrophotometer (Shimadzu, Tokyo, Japan) using KBr pallets. Elemental analyses were carried out on Heraeus Rapid Analyser (Heraeus, Germany) and functions were within 0.4% of the theoretical value.

Synthesis of 2'-chloro 4', 6'-bis-(bromo-aniline)-s-triazines

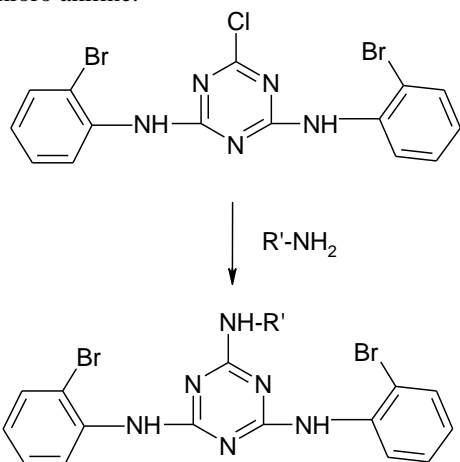
Cynuric chloride 9.3 gm was dissolved in acetone (50 ml) and cooled at 0°C to it p-bromo aniline (10 gms) was added at low temperature followed by sodium hydroxide (0.2 m) in water (25 ml) the temperature was raised to 45-50°C further stirred for 2 hours poured into ice cooled water, filtered and dried.

2.95 gm of 2'-chloro-4',6'-bis-(bromo-aniline)-s-triazine was dissolved in 50 ml of dioxane to it 1.69 gm of selected amine and sodium hydroxide (0.1 mg, 25 ml of water) were added. The constants were refluxed on the water bath for 3 hours and poured into ice cooled water.



Synthesis of 4', 6'-bis-(bromo-aniline)-2'-(arylamine)-s-triazine

The separated compound was filtered washed with water and dried the amines taken as o-toluidine, m-toluidine, p-toluidine and o-chloro aniline, m-chloro aniline, p-chloro aniline.



Com pd	AP-1	AP-2	AP-3	AP-4	AP-5	AP-6
R'	2-toluidine	3-toluidine	4-toluidine	2-chloroaniline	3-chloroaniline	4-chloroaniline

The molecular formula of these compounds were calculated from their elemental analysis and mass spectroscopy. The structure of synthesized s-triazine have been confirmed by elemental analysis, IR and ¹H-NMR spectral data studies. The yields melting points and spectral and analysis data of the synthesized s-triazines are given in tables 1 to 7

The separated compound was filtered washed with water and dried, the amines were taken as o-toluidine, m-toluidine, p-toluidine, o-chloro aniline, m-chloro aniline and p-chloro aniline.

RESULTS AND DISCUSSION

The molecular formula of these compounds was calculated from their elemental analysis and mass spectrum. The structure of synthesized s-triazine have been confirmed by elemental analysis, IR and ¹H-NMR spectral data studies. The antibacterial and antifungal activities of compounds (a to f) were assayed in vitro against selected bacteria: Escherichia coli, E. Scherichia coli, Staphylo aures, Sligelly and fungi Aspergillus niger, Aspergillus Purasities, Tricroderm uridue, Chrysoporium Sps. The minimal inhibitory concentration (MIC) values of compounds a to f were determined using the filter paper disc diffusion method (antibacterial and antifungal activity). Streptomycin and griseofulvin, used as the standard for the antibacterial and antifungal activity, respectively, showed MIC values in the range 1.25–3.25 µg mL⁻¹ for all the bacterial strain and 6.25–12.5 µg mL⁻¹ for all fungal strain. All standards were also screened under similar condition for comparison.

4', 6'- bis-(bromo-aniline)-2'-(2-toluidine)-s-triazine (AP-1) : Yield 90% , m. p. 235⁰C. Anal. Cald for C₂₂H₁₆N₆Br₂ : C 47.95 %,H 2.86 %,N 25.16 % found C 48.17 %,H 2.91 %,N 25.16 % ; IR: 2958, 1450, (-CH₃), 1556, 1417(-C=N Str.), 3020,1608,868 (Aromatic ring), 3375 (-NH str.) 1528 (NH bend.), 621 (C-Br) ; ¹HNMR: 2.56 (s, 3H, -CH₃), 6.80-7.38 (m, 12 H, Ar-H), 8.69 (s, 1H, -NH-), 8.95 (s,2H,-NH); Mass (FAB) :548 (M⁺).

4', 6'- bis-(bromo-aniline)-2'-(3-toluidine)-s-triazine (AP-2) : Yield 88% , m. p. 240⁰C. Anal. Cald for C₂₂H₁₆N₆Br₂ : C 48.05 %,H 2.77 %,N 25.15 % found C 48.17 %,H 2.91 %,N 25.16 % ; IR: 2958, 1477, (-CH₃), 1558, 1418(-C=N Str.), 3026,1602,868 (Aromatic ring), 3375 (-NH str.) 1528 (NH bend.), 590 (C-Br) ; ¹HNMR: 2.53 (s, 3H, -CH₃), 7.03-7.76 (m, 12 H, Ar-H), 8.02 (s, 1H, -NH-), 8.84 (s,2H,-NH); Mass (FAB) :548 (M⁺).

4', 6'- bis-(bromo-aniline)-2'-(4-toluidine)-s-triazine (AP-3) : Yield 86% , m. p. 243⁰C. Anal. Cald for C₂₂H₁₆N₆Br₂ : C 47.85 %,H 2.78 %,N 25.12 % found C 48.17 %,H 2.91 %,N 25.16 % ; IR: 2941, 1465, (-CH₃), 1560, 1423(-C=N Str.), 3020,1608,868 (Aromatic ring), 3375 (-NH str.) 1528 (NH bend.), 610 (C-Br) ; ¹HNMR: 2.53 (s, 3H, -CH₃), 7.03-7.76 (m, 12 H, Ar-H), 8.02 (s, 1H, -NH-), 8.84 (s,2H,-NH); Mass (FAB) :548 (M⁺).

4', 6'- bis-(bromo-aniline)-2'-(2-choloaniline)-s-triazine (AP-4) : Yield 79% , m. p. 255⁰C. Anal. Cald for C₂₀H₁₅N₆ClBr₂ : C 44.68 %,H 2.65 %,N 15.45 % found C 44.96 %,H 2.81 %,N 15.73 % ; IR: 1560, 1423(-C=N Str.), 3070,1602,933 (Aromatic ring), 3378 (-NH str.) 1510 (NH bend.), 616 (C-Br) ; ¹HNMR: 7.02-7.63 (m, 12 H, Ar-H), 8.07 (s, 1H, -NH-), 8.33 (s,2H,-NH); Mass (FAB) :533 (M⁺).

4', 6'- bis-(bromo-aniline)-2'-(3-choloaniline)-s-triazine (AP-5) : Yield 79% , m. p. 260⁰C. Anal. Cald for C₂₀H₁₅N₆ClBr₂ : C 44.62 %,H 2.69 %,N 15.48 %

found C 44.96 %,H 2.81 %,N 15.73 %; IR: 1561, 1417(-C=N Str.), 3030,1595,948 (Aromatic ring), 3378 (-NH str.), 1493 (NH bend.), 615 (C-Br); ¹HNMR: 6.98-7.90 (m, 12 H, Ar-H), 8.08 (s, 1H, -NH-), 8.25 (s,2H,-NH); Mass (FAB) :533 (M⁺).

4', 6'- bis-(bromo-aniline)-2'-(3-choloaniline)-s-triazine (AP-6) : Yield 89% , m. p. 265⁰C. Anal. Cald for C₂₀H₁₅N₆ClBr₂ : C 44.63 %,H 2.55 %,N 15.49 % found C 44.96 %,H 2.81 %,N 15.73 %; IR: 1558, 1412(-C=N Str.), 3060,1602, 989 (Aromatic ring), 3379 (-NH str.), 1519 (NH bend.), 616 (C-Br); ¹HNMR: 6.98-7.80 (m, 12 H, Ar-H), 8.01 (s, 1H, -NH-), 8.93(s,2H,-NH); Mass (FAB) :533 (M⁺).

Considerable attention has been paid to the analysis of chemical in the s-triazine group, due to their widespread use in agricultural chemistry and their subsequent degradation in biological systems. For initial chemical screening of the activity of newly synthesized compounds it is recommended first to determine their lipophilicity, an important physico-chemical property in relation to biological activity. Lipophilicity is difficult to quantitate, but the most widely accepted measure of lipophilicity is the octanol-water partition coefficient, defined as the ratio of the concentrations of the solute in the two phases of a saturated 1-octanol-water system. Measurement of the octanol-water partition coefficient is achieved by an alternative method, reversed-phase liquid chromatography. Reversed-phase thin-layer chromatography (RP TLC) has been found to offer a rapid method for the analysis of a large number of s-triazine type compounds.

Certain relationships between the structure of s-triazine compounds and their mobility on silica gel impregnated with paraffin oil have recently been demonstrated. The retention behavior of compounds in various chromatographic systems is believed to be different by nature, i.e. the different physico-chemical properties of an analyte can influence its retention. Most recently, much effort has been done with the major aim of finding a mathematical model relating the retention of a given analyte to physico-chemical and structural parameters (descriptors) of test molecules. These correlations are known as quantitative structure-retention relationships (QSRR). Besides practical application in optimization strategies, QSRR studies can significantly contribute to getting some insight into the molecular mechanism of separation. The QSRR equations describing R_{M0} determined for different mobile phase organic component in terms of logarithms of n-octanol-water partition coefficients were derived. The partition coefficients (AlogP, IAllogP, ClogP, logPKowin, XlogP, ACDlogP) were calculated by using different software packages. The purpose of the work described in this paper was, therefore, to select the logP data and TLC

system that best characterize octanol/water partitioning, and thus the Lipophilicity of the investigated molecules. Six derivative of s-triazines were investigated. Standard solutions (1 mg cm⁻³) were prepared in methanol, acetone, or chloroform. Samples were spotted on the plates by means of a micro-pipette. TLC was performed on 20 × 20 cm glass plates precoated with impregnate silica gel. The thin-layer of impregnate silica gel was prepared by suspending 25 g silica gel 60 GF254 (Merck) in 100 ml diethyl ether containing 2.5 % paraffin oil. To ease the visualization, fluorescent indicator F254 (Merck) was incorporated into the layers [10]. Impregnate silica gel layer was developed using the following mobile phases: Aprotic solvents: Acetonitrile-water (φ=0.2-0.6; v/v), Acetone-water (φ= 0.5-0.8; v/v), Tetrahydrofuran-water (φ=0.45-0.7; v/v), Dioxane-water (φ =0.5-0.8; v/v). Protic solvents: Methanol-water (φ=0.5-0.8; v/v), Ethanol-water (φ =0.5-0.8; v/v).The plates were developed to a distance of 15 cm by the ascending technique at room temperature without previous saturation of the chamber with mobile phase. Dark spots were observed under UV light (λ =254 nm). R_M values were calculated from $R_M = \log[(1/R_f) - 1]$. All calculations were performed using the computer software Origin, Version 6.1. The partition coefficients AlogP, IAllogP, ClogP, logP_{Kowin} and XlogP, were calculated for the compounds by applying different theoretical procedures [11, 12]. ACDlogP was calculated using commercial software and the other partition coefficients were obtained from the internet [13].

Determination of Retention Constants, R_{M0} , TLC Equations:

When the R_M values calculated from R_f values (retention factor defined as the distance migrated by the sample from the origin compared with the distance migrated by the solvent front from the origin) were plotted against mobile phase composition for each compound there was a range in which a linear relationship was observed between the R_M values and organic modifier concentration in the mobile phase, which can be expressed by the equation $R_M = R_{M0} S\phi$, indicative of the reversed-phase chromatography, where φ is the amount (%) of organic compounds in the mobile phase [14]. The obtained slopes, S, and intercept values, R_{M0}, of TLC equation for each solute are presented in Table 1. The correlation coefficients of the TLC equations were satisfactory.

Correlation Between Retention Constants, R_{M0}, and Slope, S:

A linear relationship was observed between the intercept, R_{M0}, and slope, S, for protic and aprotic solvents, as shown by the equations given in Table 3.3.3.

The best correlation was obtained for acetone as mobile-phase modifier (r = 0.994). There is a good correlation between R_{M0} and S, which might reflect the suitability

of the systems examined for estimating the lipophilicity of the compounds. The RM_0 values, which are chromatographic data describing the partitioning between a non-polar stationary and a polar mobile phase, may therefore be appropriate for the assessment of lipophilicity.

Table:1: Equation for the relationships between the retention constant R_{M0} and Slope, S

Mobile Phase	Equation	r	SD	n
Acetone-Water	$R_{M0} = - 0.572 - 1.121S$	0.994	0.101	18
Acetonitrile-Water	$R_{M0} = -1.717 - 1.356S$	0.954	0.590	18
Dioxane-Water	$R_{M0} = - 1.895 - 0.965S$	0.920	0.392	18
Tetrahydrofuran-Water	$R_{M0} = - 1.895 - 0.965S$	0.985	0.140	18
Methanol-Water	$R_{M0} = - 0.231 - 1.231S$	0.987	0.110	18
Ethanol-Water	$R_{M0} = - 0.786 - 1.152S$	0.962	0.283	18

Based on the results obtained on silica gel impregnated paraffin oil, RM_0 is directly dependent on the nature of mobile phase modifiers. In other words, the selectivity of separation of the tested substances are the result of specific interactions with the mobile phase.

Correlation of Retention Constants, RM_0 and logP:

Lipophilic character often seems to be the most important physico-chemical parameter in determining the biological activity of chemical agents. Lipophilicity can be expressed in terms of many different descriptors (logP, logkw, RM, RM_0), obtained experimentally or calculated. The experimental parameters most frequently used are the retention constants RM_0 (RPTLC) and logkw (RPHPLC), whereas the calculated quantity is logP. The partition coefficient, logP, of a given compound between a non-aqueous and an aqueous phase can be used as an expression of its lipophilic character [15]. Because the retention of a compound in reversed-phase chromatography is governed by hydrophobic interactions, linear relationships between the retention constant, RM_0 , and logP could be expected [16]. The partition coefficients (AlogP, IAllogP, ClogP, logPKowin, XlogP, ACDlogP) of striazine derivatives are listed in **Table: 1**

Table:2:. Partition coefficients calculated by different theoretical methods

Compd	AlogP	IAllogP	ClogP	logP _{Kowin}	XlogP	ACDlogP
AP-1	6.18	5.89	6.58	6.85	6.43	5.42
AP-2	6.22	5.91	6.66	6.89	6.41	5.49
AP-3	6.16	5.93	6.61	6.88	6.46	5.45
AP-4	5.55	4.81	5.48	6.86	5.14	4.82
AP-5	5.58	4.86	5.49	6.81	5.19	4.86
AP-6	5.59	4.88	5.46	6.84	5.15	4.86

Table: 2:. Correlation coefficients (r) for the correlation between RM_0 and different logP values

Compd	AlogP	IAllogP	ClogP	logP _{Kowin}	XlogP	ACDlogP
Acetone	0.835	0.705	0.858	0.877	0.795	0.804
Acetonitrile	0.763	0.708	0.828	0.875	0.671	0.757
Dioxane	0.747	0.624	0.815	0.900	0.681	0.770
Tetrahydrofuran	0.570	0.533	0.648	0.756	0.532	0.707
Methanol	0.702	0.604	0.794	0.895	0.665	0.750
Ethanol	0.855	0.795	0.822	0.838	0.900	0.825

By comparing the calculated values to define the lipophilicity of the investigated molecules, it is evident that ethanol as a modifier gives the highest correlation (calculated average correlation coefficient is 0.839). Retention Constants, RM_0 for QSRR. The QSRRs are statistical models which quantify the relationship between the structure of a molecule and its chromatographic retention parameters in different kinds of chromatography. The application of QSRR allows the prediction of the retention of a new solute, identification of the most informative structural descriptors, elucidation of the molecular mechanisms of separation in a given chromatographic system, evaluation of complex physico-chemical properties of solutes and estimation of biological activities.

The relationship between the retention and the structural characteristics of a molecule explains the effect of chemical structure on the retention behavior in a more accurate way. The use of multiple linear regression (MLR) analysis for fourteen s-triazine derivatives led to statistically significant equations relating lipophilicity (estimated by R_{M0} values (dependent variable) to different theoretically calculated six types of log P namely AlogP, ClogP, ACDlogP, logPKowin, X logP and IAllogP values for each compound (independent variable). The specifications for the best-selected MLR models are shown in Table 3 and Table 4.

These relationships were analyzed and the best model was selected on the basis of various statistical parameters like correlation coefficient (r), and standard deviation (SD). In the first phase of work, the multilinear relationships between the retention constant and two variable lipophilicity descriptors was examined.

Table: 3 :: Statistical parameters for multilinear dependence between R_{M0} and two variables descriptors Lipophilicity

Modifier	Descriptors		$R_{M0} = a \log p_1 + b \log p_2 + c$				
	$\log p_1$	$\log p_2$	A	b	c	r	SD
Acetone	$\log P_{Kowin}$	XlogP	0.0993	0.648	-0.313	0.916	0.329
Dioxane	AlogP	$\log P_{Kowin}$	0.312	0.813	-0.410	0.949	0.306
Methanol	$\log P_{Kowin}$	XlogP	0.571	0.672	-0.357	0.953	0.234

The analysis of these results indicates that the proposed models can correctly represent the relationship between the retention parameters of the investigated compounds on silica gel and different log P values calculated for various compound solely from the molecular structure. These models are suitable for prediction of the retention of structurally similar compounds under the same chromatographic conditions.

Table: 4: Statistical parameters for multilinear dependence between R_{M0} and three variables descriptors Lipophilicity

Modifier	Descriptors			$R_{M0} = a \log p_1 + b \log p_2 + c \log p_3 + d$					
	$\log p_1$	$\log p_2$	$\log p_3$	A	b	c	d	r	SD
Acetone	AlogP	$\log P_{Kowin}$	XlogP	-1.969	0.811	0.524	-0.632	0.948	0.274
Acetonitrile	AlogP	$\log P_{Kowin}$	XlogP	0.731	0.853	-0.292	-0.231	0.957	0.294
Dioxane	$\log P_{Kowin}$	XlogP	IAllogP	-2.713	1.401	0.756	-1.203	0.901	0.659
Tetrahydrofuran	$\log P_{Kowin}$	XlogP	ACDlogP	0.664	0.271	-0.609	0.791	0.904	0.280
Methanol	$\log P_{Kowin}$	XlogP	IAllogP	0.948	0.708	-0.251	-0.208	0.964	0.216
Ethanol	$\log P_{Kowin}$	XlogP	ClogP	0.625	0.408	0.537	-0.503	0.907	0.398

Experimentally obtained R_f R_{M0} values depend on the nature of organic modifier in the mobile phase. A linear relationship between R_{M0} and slope, S, values was found for all mobile phases. Satisfactory linear correlation was obtained between the retention constants and AlogP, ClogP, ACDlogP, logPKowin, X logP and IAllogP. According to the correlation coefficients, R_{M0} is a useful property for evaluation of the relative lipophilicity of the examined compounds. The correlations between the retention constants, R_{M0} , and selected lipophilicity parameter (different logP values) of the solutes were expressed by multiparametric equations of high statistical significance, indicate that these models can be used to predict the retention constants of these molecules.

Table- 5: Antibacterial activity of the newly synthesized derivatives

Compound code	Escherichia coli		Bucillus Subsniss		Staphyloi aures		Sligelly	
	2%	4%	2%	4%	2%	4%	2%	4%
AP-1	++	+++	++	+	++	+++	+	++
AP-2	++	-	++	+	++	+++	+	+++
AP-3	+	-	++	++	+	+++	+	+++
AP-4	+	++	+	++	++	+++	++	++
AP-5	++	+++	++	+++	++	+++	++	+++
AP-6	++	++	++	++	++	+	++	++

Std- = streptomycin inhibition diameter in mm

Highly active = +++ (inhibition zones >15)
 moderately active = ++ (inhibition zone 10-15)
 slightly active = + (inhibition 10)
 inactive inhibition zone < -6) For bacteria

Table 6 :Antifungal activity of the synthesized compound derivatives

Compound code	<i>Aspergillus niger</i>		<i>Aspergillus Purasities</i>		<i>Tricroderm uridue</i>		<i>Chrysoporium Sps</i>	
	2%	4%	2%	4%	2%	4%	2%	4%
AP-1	++	+++	++	+	++	++	++	+++
AP-2	++	+++	++	+++	++	++	+	++
AP-3	++	++	+	++	+	++	+	++
AP-4	++	++	+	++	+	+	+	++
AP-5	-	++	-	++	-	++	-	+++
AP-6	+	+++	-	++	-	++	-	+++

Std- Exriseofulvin inhibition diameter in mm
 Highly active = +++ (inhibition zones >15)
 moderately active = ++ (inhibition zone 10-15)
 slightly active = + {inhibition 10}
 Inactive = - (inhibition zone) Zone < -6) for bacteria

RESULTS AND DISCUSSION

Given rise to the the final product a to f most of the compound of the series where white crystalline which where systematically crystallize and purified before sent for elemental analysis and there structural characterization. The Synthesised compounds where simulated on the computer using PC Model Software [Sarena Version 3.01] visualize steric arrangement and the spatial elongation of the substitutes groups and they relative position. As expected form the molecular structure and their substitution.

Table-7: Variation of IR, ¹HNMR and PC Model data values for different substituted derivatives

Compd	Dipole Moment	VdW Force	Molar Volume	IR cm ⁻¹		¹ HNMR ppm		
				C-N str	N-H str	-NH	-NH	Ar-H (C-H)
AP-1	2.660	16.660	277	1417.7	3370.0	8.35	8.72	6.96-7.89
AP-2	2.183	16.183	277	1418.2	3377.0	8.32	8.54	6.99-7.69
AP-3	2.591	16.591	277	1415.3	3377.3	8.22	8.36	7.12-7.63
AP-4	1.052	16.152	273	1423.8	3378.0	8.15	8.30	7.10-7.72
AP-5	2.748	16.748	273	1417.7	3377.5	8.16	8.25	7.05-7.76
AP-6	2.917	16.917	273	1412.2	3379.5	8.41	8.87	7.01-7.73

Thus an overall comparison of the compounds in this series viz. there overall steric performance relative variation in the simulation data and the values obtained and result of characterization justifies the selection of these compounds for synthesis in the present work.

ACKNOWLEDGEMENT

The authors are thankful to SAIF, Central Drugs Research Institute Lucknow (India) for providing the spectral and analytical data of the compounds. We are thankful to Head Department of Chemistry Govt. p. g. college Chhindwara for providing the facilities to carry out the work and the Head, Department of Microbiology UTD sagar for the microbial activity. We are also thankful to the Dr.(Mrs.) Sandhaya Shrivastava, Head,

Department of Chemistry, Govt. P.G. College Seoni M.P., (India) for his suggestions and guidance in the research work

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