

DENSITY OF COLIFORM BACTERIA IN RIVER NARMADA AT OMKARESHWAR CITY, MADHYA PRADESH, INDIA

1.Priyanka Laad, 2.C.S. Shrivastava

1. Department of Zoology, Govt. S.B.N.P.G. College Barwani MP- 451551.
2. Department of Zoology, Govt Holkar Science College Indore MP- 452001

ABSTRACT: Water resources are being polluted due to increasing urbanization, modern practices in agriculture and other human activities. The link between human health and environmental pollution is of significant dimension. Studies were conducted with an objective of assessing the coliforms density in river Narmada water. River is polluted with domestic and agriculture wastes of Omkareshwar city. Water samples were collected from five designated sampling stations of the river. Results revealed that, all the samples from the selected stations contained coliforms bacteria above the safe limits for drinking water standards. Seasonal variations in coliforms density were recorded with maximum density in summer and monsoon seasons and minimum density in winter seasons.

KEYWORDS: Coliform bacteria, Water, River, Pollution

INTRODUCTION

Water contamination by various routes and agents are liable to cause clinically signified outbreaks of intestinal infections in both animals and man. The demonstration of pathogenic bacteria, would obviously constitute the most direct proof of a hazardous impurity; but these pathogens, if present are so scanty that the technical difficulty of their isolation makes the test impractical for ordinary purposes and they are very sensitive even to a slight change of environmental factors and difficult to isolate. Hence, it is impractical to attempt directly to detect the presence of all the different kinds of water borne pathogens, any of which may be present only intermittently. Thus, we rely on tests that reflect the presence of commensal bacteria of intestinal origin such as those of the coliform group, which are more numerous, more easily tested and are the most reliable indicators of fecal pollution (Senior 1989). Each person discharges 100 to 400 billion coliform organisms/day, in addition to other kinds of bacteria. These coliforms do not themselves constitute a hazard since, they are harmless to man and are in fact useful in destroying

organic matter in biological waste treatment processes. But they indicate that faecal matter has entered and that the water is therefore, liable to contamination with more dangerous organisms (Senior 1989). Presence of coliform organisms is thus taken to indicate possible presence of pathogenic organisms. Coliform monitoring being rapid, inexpensive easy to perform, proved effective (Leclerc *et al.*, 2000). Coliform was the term first used in 1880s to describe rod-shaped bacteria. Coliform group of bacteria is functionally-related groups belong to a single taxonomic family Enterobacteriaceae and comprises many genera and species. The term coliform organisms refers to gram-negative, rodshaped bacteria capable of grow in presence of bile salts or other surface-active agents with similar growth-inhibiting properties and able to ferment lactose at 35°C-37°C with the production of acid, gas and aldehyde within 24-48 hours (Clesceri *et al.*, 1998). They are also oxidase-negative and non-spore-forming and display β-galactosidase activity (Arora 2003).

Traditionally coliform bacteria were regarded as belonging to the genera *Escherichia*, *Citrobacter*, *Enterobacter* and *Klebsiella*. MacConkey (1909) defined coliform types and this number was increased to 256 by a system developed by Bergey and Deehan (1908). However, as defined by modern taxonomic methods, the group is heterogenous consisting of as many as 32 genera (Leclerc *et al.*, 2001). Some of these can be found in both faeces and the environment (nutrient-rich waters, soil, decaying plant material) as well as in drinking water with relatively high concentration of nutrients and may multiply. Eg. *Serratia fonticola*, *Rahnella aquatilis*, *Buttiauxella agrestis*. Coliform bacteria can be indicators for potential pathogens responsible for various waterborne diseases (Hunter *et al.*, 2004) or be pathogens themselves, such as certain strains of *E. coli*. In the present experiment we tested water samples of Narmada River for abundance and species richness of coliform and other environmental bacteria. We hypothesized that river with more human or animal anthropogenic activities or watershed with agricultural or sewage runoff would have higher bacterial abundance and species richness.

MATERIAL AND METHODS

The present study was of the river Narmada at Omkareshwar dam reservoir near Omkareshwar city. Water samples from Narmada river were collected during August 2011 to July 2013 (24 months) from all the five designated stations and were analysed for the levels (MPNs) of generally accepted faecal pollution indicators i.e., Total coliforms (TC) and Faecal coliforms (FC). Seasonal variations were examined among these indicators during August 2011 to July 2013 as well as the species composition for each group of indicators. The pollution range, maxima, minima, standard deviation of the above parameter has been given along with the detailed discussion of the parameter results. Presumptive coliform count: Single and Double strength MacConkey's broth medium (Hi-media, Ltd.) was prepared by dissolving 4 g and 8 g of MCB in 100 ml of distilled waters separately. Measured amounts were sterilized in test tubes containing a Durham's tube for indicating gas production. With sterile graduated pipettes or with adjustable micropipettes that can hold sterilized microtips, the following amounts of the sampled waters were added.

- a. Five 10 ml quantities each to DS (Double Strength) medium.
- b. Five 1 ml quantities each to 10 ml SS (Single strength) medium.
- c. Five 0.1 ml quantities each to 10 ml SS medium.

Tubes that show acid and sufficient gas to fill the apex covering the concavity of Durhams tube at 24 hours were considered to be presumptive positive, as a result of the growth of coliform Bacilli after incubation at 37°C. Incubation was sustained from the next session; tubes showing presumptive positive at 48 hours of incubation were accredited to the existing numbers in the concerned station. Sample dilutions were read separately and the positives in A, B and C were serially collated in probability tables to obtain presumptive coliform Bacilli in 100 ml of the sampled station. Tubes showing gas production at 37°C for 48 hours were confirmed by plating in MacConkey agar plates (MCA) and examined for typical colonies.

RESULTS AND DISCUSSION

In the present investigation was selected stations (S I, S II, S III, S IV and S V) as first independent variable and as second independent variable against all parameters considered as dependent variables. The maximum, minimum and averages of two years in the Total Coliforms and Faecal Coliforms are represented in table-

1 to table-4. Presence of coliform organisms in water has been attributed to influxes of allochthonous bacteria from waste discharges and surface water drainage. Total coliform levels were always higher than faecal coliform levels, which is not surprising since total coliforms can originate from non-faecal sources also such as plants and soils (Geldreich, 1974). The above findings and opinions also were corroborated by Goyal *et al.* (1977).

In the present study, high coliform load has been registered during the entire course of study. The higher values of MPN for coliforms were recorded at S II and S III, mainly attributed to the domestic sewage and municipal wastes. Though, the values remain statistically significant. Higher coliform levels occurring during summer months suggest survival and high influx of the coliforms and also increased nutrient influx from decomposition of summer productivity because of high temperature and watershed drainage may also contribute to March-April increase in numbers of both the years. In addition, heavy usage of the river was found during summer for bathing, washing and such other anthropogenic activities, that also would contribute for higher incidence of coliforms. Water quality of Narmada river showed significant deterioration in view of global standards. All most all the samples were contaminated with coliform bacteria, resulting mainly due to anthropogenic activities, especially discharging of domestic and agricultural wastes directly into the river as some recent studies (Sharma *et al.*, 2010; Sadat *et al.*, 2011) revealed that coliform count has positive relation with anthropogenic activities. Present results are in agreement with the findings of Geldrich (1972) who observed an increase in the total coliform count of water bodies due to increased use of animal wastes as manure in the agricultural fields.

In contrast, Goyal *et al.*, (1977), Sayler *et al.*, (1975) found higher organisms in winter months than in summer. In the present study winter maxima was followed by rainy season. Feachem (1974), Gerba and Schaiberger (1975) have also reported an increase in the number of indicator organisms and Faechem (1974) found peaks of coliforms during on immediately after a storm and other times due to unexplained factors. Occurrence of coliforms in the samples is also confirmed by study conducted by Lateef *et al.*, (2003) reporting high coliform counts in the spring water of Kashmir valley. Due to change in agricultural methods, diminishing of livestock farms, intensive farming operation increased concentration of animal wastes results in an increased pollution of rivers and streams (Gelt1998). High coliform incidence is influenced by the entry of surface runoff water from nearby areas that carried higher levels of suspended matter and nutrients

on their surface. Similar observations were made by Venkateswarlu (1986) in the rivers of Andhra Pradesh. According to Keller (1960) access of soil and organic matter and increase in temperature is responsible for the increase in bacterial numbers in river water after rain and during summer. This is supported by Raj Kumari Sing *et al.* (2001) who carried investigations in Narmada River at Hoshangabad. In Lake Windermere, Taylor (1940) has observed that there is a close relationship between the bacterial content of the water and the rainfall in the drainage area during the preceding week. Subramanyan and Bhaskaran (1948), Voelkar *et al.* (1960) noted a considerable decrease in the number of coliforms in dug wells during cold months. Further, S II and S III have bathing ghats, where disturbance due to human activities is seen throughout the year. Highest incidence was recorded in S IV followed by S III, S II and S I. Coliform load in river water indicated that bacteria were always and under all conditions remains in water body (Agarwal 1993). Studied of Prathibha and Murulidhar (2015) High counts of bacteria signified the organic pollution in river water of Tunga at shivamogga city, Karnataka, India. Extremely high MPN values of Total coliforms i.e., 1600x1010 were reported near Opium factory in Ganga water (Shukla Suresh *et al.*, 1992). They also quote the observations of other workers that there is a direct effect of sewage containing faecal matter on the growth and population of coliform bacteria. Agarwal (1993) registered a range of 19-22000/100 ml of TC in river Betwa. Kataria *et al.*, (1997) studied river Halali and reported 240 to 2400/100 ml. Doctor *et al.*, (1998) recorded MPN between 300 to 1600/100 ml in river Bhadar. Raka *et al.*, (1999) registered the faecal pollution in drinking water of Maharashtra and reported the coliform in range of 21 180/100 ml. Koshy Mathew and Vasudevan Nayar (2000) point out that, MPN of Pamba river water was infinite at four of the seven stations considered due to Sabarimala pilgrimage and free flow of sewage, domestic waste and faecal matters into the river at those stations.

Table.1 Sectoral range and mean values of total coliforms (TC/100ml) from August 2011 to July 2012.

Sampling Stations	minimum	maximum	Mean	Standard Deviation
S-I	38	120	54.66	21.58
S-II	39	130	59.33	24.16
S-III	32	122	58.08	23.80
S-IV	45	110	59.17	18.02
S-V	29	100	41.67	19.28

Table.2 Sectoral range and mean values of total coliforms (TC/100ml) from August 2012 to July 2013.

Sampling Stations	minimum	maximum	mean	Standard Deviation
S-I	47	132	61.75	23.09
S-II	42	132	61.42	24.24
S-III	30	134	61.83	26.19
S-IV	51	120	69.58	18.28
S-V	35	95	45.42	16.25

Table.3 Sectoral range and mean values of faecal coliforms (FC/100ml) from August 2011 to July 2012.

Sampling Stations	minimum	maximum	mean	Standard Deviation
S-I	07	18	14.83	8.96
S-II	10	39	18.25	7.84
S-III	15	31	22.58	4.68
S-IV	16	42	28.83	8.52
S-V	06	24	13.92	5.33

Table.4 Sectoral range and mean values of faecal coliforms (FC/100ml) from August 2012 to July 2013.

Sampling Stations	minimum	maximum	mean	Standard Deviation
S-I	10	19	17.66	8.74
S-II	14	33	22.5	5.76
S-III	18	32	26.25	4.22
S-IV	25	44	32	5.51
S-V	13	29	19.5	4.83

They also indicate an increased coliform content at the sides of the Pampa River (1600/100 ml) and very low values at the middle part (12/100 ml). They attribute this to the good velocity at middle with no flow near the sides due to sand mining. Shafi *et al.*, (2013) observed that, all the samples obtained from the Manasbal Lake of Kashmir were positive with respect to coliform occurrence, though the count was variable ranging between 4 and 460 MPN/100 ml highest proportion of indicator coliforms was found in water samples collected at the site surrounded by residential hamlets in comparison to other three sites. Debashish Mandal and Mukhyopadhyay (1991) also report extremely high MPN ranging from 10x102 to 9x106/100 ml in river Ganga. According to them coliform bacteria are more abundant in certain sewage channels and bathing ghats. Musaddiq Mohammed (2000) recorded a maximum MPN of 900 in

Morna River and suggests health education as an important weapon in creating desire for high standard of life. Rajkumar Singh *et al.*, (2001) noted MPN index up to 9200/100 ml in Narmada River signifying the organic pollution in the water body due to the human activities. Asthana and Singh (1993) observed extremely high MPN values of 21×10^3 to 40×10^3 in Gomati River with faecal coliform accounting for nearly 60 to 80% of the total coliform. Praveen Jain and Sanjay Telang (1996) have shown an MPN of 118 in Parbathy River at Sehore (M.P) and conclude that river water has minimal bacterial load. Dutka (1973), Evison and James (1973), Burrel and Roland (1979), Hazen (1988), Ramteke *et al.*, (1992) advocate that, total coliform count performed by the most probable number (MPN) method is commonly used indicator of potability despite the drawbacks that have been well documented.

CONCLUSION

It is concluded from the present results that, existence of both non-faecal bacteria that fit the definitions of coliform bacteria and of lactose-negative coliform bacteria limits the applicability of this group as an indicator of faecal pollution. Drinking directly from river without filtration should be avoided at all times. Many organisms present in river are very sensitive to ecosystem. Coliform bacteria should not be detectable in treated water supplied and if found, suggest inadequate treatment, post treatment contamination or excessive nutrients. Coliform test can therefore be used as indicator for both treatment efficiency and integrity of distribution system. Although coliform organisms may not always be directly related to the presence of faecal contamination or pathogens in drinking water, coliform test is still useful for monitoring the microbial quality of treated piped water supplies. If there is any doubt, especially when coliform organisms are found in the absence of thermo tolerant coliforms and *E. coli*, identification to the species level or analyses for other indicator organisms may be undertaken to investigate the nature of contamination. Sanitary inspections will also be needed.

REFERENCES

1. Agarwal, A. 1993. Studies on physicochemical and biological characteristics of river Betwa from Nayapura to Vidisha. Ph.D. Thesis, Barkatulla University, Bhopal.
2. Arora, D.R. 2003. Text book of microbiology, 2nd edn. CBS publishers and distributors, New Delhi. Pp. 653-659.
3. Asthana, R.K., Singh, K.N. 1993. Physicochemical characterization of Gomathi River. *Oriental J. Chem.*, 9(2):155-157.
4. Bergey, D.H., Deehan, S.J. 1908. The colon aerogenes group of bacteria. *J. Med. Res.*, 19: 175.
5. Burrel, R.A., Roland, M.G. 1979. The relationship between rain fall and well water pollution in a West African (Gambia) village. *J. Hygiene.*, 83: 143-150.
6. Clesceri, L.S., Greenberg, A.E., Eaton, A.D. 1998. Standard Methods for the Examination of Water and Wastewater, 20th ed., APHA, Washington, D.C. Pp.13-25.
7. Debashish Mandal and Mukhyopadhyay, S. 1991. Incidence of Coliphages in the Ganga water and its sewage out-falls around Calcutta-an approach towards biological monitoring of water pollution. *Sci. Cult.*, 57(8&9): 20-23.
8. Doctor, P.B., Raiyani, C.V., Verma, Y., Desai, N.M., Kulkarni, P.K., Ruparelia, S.G., Ghosh, S.K. 1998. Physicochemical and microbial analysis of Dye contaminated river water. *Indian J. Environ. Hlth.*, 40(1): 7-14.
9. Dutka, B.J. 1973. Coliforms are inadequate index of water quality. *J. Environ. Hlth.*, 36: 39-46.
10. Evison, L.M., James, A. 1973. A comparison of the distribution of intestinal bacterial in British and East African water sources. *J. Appl. Bacteriol.*, 36: 109-118.
11. Faechem, R. 1974. Faecal coliforms and faecal *Streptococci* in streams in the New Guinea Highlands. *Water Res.*, 8: 367-374.
12. Geldreich, E.E. 1972. Buffalo lake recreational water quality: a study in bacteriological data interpretation. *Water Res.*, 6: 913-924.
13. Geldreich, E.E. 1974. Microbiological criteria concepts for coastal bathing waters. *Ocean Manage.*, 3: 225-248.
14. Gelt, J. 1998. Microbes increasingly viewed as water quality threat. *Arroyo.*, 10:1.
15. Gerba, C.P., Schaiberger, G.E. 1975. Effect of particulates on virus survival in seawater. *J. Water poll. Control Fed.*, 47: 93-103.
16. Goyal, S.M., Gerba, C.P., Melnick, J.L. 1977. Occurrence and distribution of bacterial indicators and pathogens in canal communities along the Texas coast. *Appl. Environ. Microbiol.*, 34(2): 139-149.
17. Hazen, T.C. 1988. Faecal coliforms an indicator in tropical water: A review. *Toxicity Assess.*, 3: 461-477.
18. Hunter, A.J., Northup, D.E., Dahm, C.N., Boston, P.J. 2004. Persistent coliform contamination in

- Lechuguilla cave pools. *J. Cave Karst Stud.*, 66: 102-110.
19. Kataria, H.C., Iqbal, S.A., Shandilya, A.K. 1997. MPN of total coliform as pollution indicator in Halali River water of Madhya Pradesh. *Poll. Res.*, 16(4): 255 -257.
20. Keller, P. 1960. Bacteriological aspects of pollution in the Juskat / Suskel Crocodile river system in Transval, South Africa. *Hydrobiologia*, 14(3-4):205-254.
21. Koshy Mathew and VasudevanNayar, T. 2000. Water quality of river Pambar at Kozencherry. *Poll. Res.*, 19(4): 665-668.
22. Lateef I, Manzoor A, Thoker and Yousuf, A.R. 2003. Bacteriological survey of 15 springs of Kashmir. *J. Res. Dev.*, 3.
23. Lateef, A., Oloke, J.K., Gueguim Kana, E.B. 2005. The Prevalence of bacterial resistance in clinical, food, water and some environmental samples in Southwest Nigeria. *Environ. Monit. Assess.*, 100: 59-69.
24. Leclerc, H., Edberg, S.C., Pierozo, V., Delattre, J.M. 2000. Bacteriophages as indicators of enteric viruses and public health risk in ground waters. *J. Appl. Microbiol.*, 88: 5-21.
25. Leclerc, H., Mossel, D.A.A., Edgerb, S.C., Struijk, C.B. 2001. Advances in the bacteriology of coliform group: Their suitability as markers of microbial water safety. *Ann. Rev. Microbiol.*, 55: 210-234.
26. MacConkey, A. 1909. Further observations on the differentiation of lactose fermenting Bacilli with special reference to those of intestinal origin. *J. Hyg.*, 9: 86-103.
27. Musaddiq Mohammed. 2000. Surface water quality of Morna River of Akola. *Poll. Res.*, 19(4): 685-691.
28. Praveen Jain, Sanjay Telang, 1996. Physicochemical analysis of water of Parbati River of district Sehore (Madhya Pradesh). *Orient J. Chem.*, 12(1): 97-100.
29. Prathibha S. and Murulidhar V.N. 2015. Diversity and Density of Coliform Bacteria in River Tunga at Shivamogga city, Karnataka, India. *Int. J. Curr. Microbiol. App. Sci.* 4 (7) 624-631.
30. Raj Kumari Singh, Iqbal, S.A., Seth, P.C. 2001. Bacteriological pollution in a stretch of river Narmada at Hoshangabad, Madhya Pradesh. *Poll. Res.*, 20(2): 211-213.
31. Raka, V.K., Agnihotri, A.R., Thekdi, R.J., Shikrolakar, S.B., Salunke, S. 1999. Efficacy of rapid field test to detect faecal pollution in drinking water. *Poll. Res.*, 18(1): 37-42.
32. Ramateke, P.W., Bhattacharjee, J.W., Pathak, S.P., Kalra, N. 1992. Evaluation of coliforms as indicators of water quality in India. *J. Appl. Bacteriol.*, 72: 352- 356.
33. Sadat, A., Akaki, K.D., Ngoran, E.B.Z., Parinet, B., Frere, J. 2011. Evaluation of Bacteriological Pollution of Yamoussoukro Lakes (Côte D ivoire). *C. Res. J. Bio. Sci.*, 3(4): 318-321.
34. Saylor, G.S., Nelson, N.D., Justice, A. Jr., Colwell, R.R. 1975. Distribution and significance of faecal indicator organisms in the upper Chesapeake Bay. *Appl. Microbiol.*, 30(4): 625-638.
35. Senior, B.W. 1989. Examination of water, milk, food and air. In: Collee, J.G., Duguid, J.P., Fraser, A.G., Marmion, B.P. (Eds), Mackie and McCartney practical medical microbiology. Churchill Livingstone, Edinburgh. Pp. 603-609.
36. Shafi, S., Kamili, A.N., Manzoor A. Shah, Bandh, S.A. 2013. Coliform bacterial estimation: A tool for assessing water quality of Manasbal Lake of Kashmir, Himalaya. *Afr. J. Microbiol. Res.*, 7(31): 3996-4000.
37. Sharma, P., Sood, A., Sharma, S., Bisht, S., Kumar, V., Pandey, P., Gusain, M.P., Gusain, O.P. 2010. Bacterial indicators of faecal pollution and physiochemical assessment of important North Indian lakes. *RMZ-Mate. Geoenviron.*, 57(1): 25-40.
38. Shukla Suresh, C., Tripathi, B.D., Mishra, B.P., Chaturvedi, S.S. 1992. Physicochemical and bacteriological properties of the water of river Ganga at Ghazipur. *Comp. Phys. Eco.*, 17(3): 92-96.
39. Subrahmanyam, K., Bhaskaran, T.R. 1948. Water-supply systems in rural areas of India: West Bengal. *Indian J. Med. Res.*, 36: 211.
40. Taylor, E.W. 1940. The examination of waters and water supplies (Thresh, Beale and Suckling) J&A. Churchill Ltd., London.
41. Venkateshwaralu, V. 1986. Ecological studies on the rivers of Andhra Pradesh with special reference to water quality and pollution. *Proc. Indian Sci. Acad.*, 96(6):495-508.
42. Voelker, R.A., Heukelekian, H., Orford, H.E. Dr. Eng. 1960. Seasonal coliform variations in well waters. *Am. J. Public Health*, 50(12): 1873-1881.